

Central University of Punjab



M.Sc. Bioinformatics

Batch 2022

Department of Computational Sciences

School of Basic Sciences

Graduate Attributes

In line with the syllabus of M.Sc. Bioinformatics it is expected that a student graduating after successful completion of the course shall be able to

- Understand the broad base of knowledge in information technology, mathematics and biology and use biological data generation technologies.
- Apply bioinformatics tools and techniques to analyse and interpret biological data.
- Build, simulate and interpret dynamic models of complex biological systems furthering basic and applied research.
- Solve the problems in real healthcare or industry scenarios using bioinformatics and computational analysis.
- Develop interpersonal skills to work as a team on a multidisciplinary front to accomplish a common goal.
- Use the knowledge, general competence, and analytical skills on an advanced level needed in the industry, consultancy, education, research, or public administration.
- Continue life-long learning as an autonomous learner and apply and nurture critical and creative thinking.

Course Structure of the Programme

SEMESTER I							
S. No.	Course Code	Course Title	Course Type				
				L	T	P	Cr
1	BIM.506	Biological Sciences	CC	3	0	0	3
2	BIM.507	Sequence Analysis	CC	3	0	0	3
3	BIM.508	Structural Bioinformatics	CF	2	0	2	3
4	PCP.519	Python Programming	CC	3	0	0	3
5	BIM.509	Genomics and Proteomics	CC	3	0	0	3
6	BIM.510	Genomics and Proteomics Lab (Practical)	SBC	0	0	4	2
Choose any one of these courses							
7	BIM.511	Protein Engineering	DE	3	0	0	3
8	BCH.511 OR HGE.528	Genetics OR Population Genetics and Genetic Epidemiology	DE	3	0	0	3
9	BOT.554	Evolutionary Biology	DE	3	0	0	3
10	BCH.512	Animal Cell Culture Technology	DE	3	0	0	3
11	ZOL.525	Nanobiology	DE	3	0	0	3
12	BIM.529	Linux and Perl	DE	3	0	0	3
Interdisciplinary course offered from other departments							
13	BIM.513	Introduction to Structural Biology	IDC	2	0	0	2
Total				19	0	6	22

SEMESTER II							
S. No.	Course Code	Course Title	Course Type				
				L	T	P	Cr
1	BIM.514	Biomolecular Simulations and Drug Design	CC	3	0	0	3
2	BIM.515	Data Mining and Machine learning	CC	3	0	0	3
3	BIM.516	NGS sequence data analysis	CF	3	0	0	3
4	BIM.517	Biostatistics using R	CC	3	0	0	3
5	BIM.518	Data Mining and Machine learning Lab (Practical)	SBC	0	0	4	2
6	BIM.519	Biostatistics using R Lab (Practical)	SBC	0	0	4	2
7	BIM.520	Bioinformatics for Next Generation Sequencing Data	VAC	2	0	0	2
Choose any two of these courses							
8	BIM.521	Big Data Analytics in Healthcare	DE	3	0	0	3
9	BIM.522	Cheminformatics	DE	3	0	0	3
10	BIM.523	Molecular Evolution	DE	3	0	0	3
10	CCC.525	Computational Methods	DE	3	0	0	3
11	LMM.525	Stem Cell and Regenerative Medicine	DE	3	0	0	3
12	MIC.556	Genetic Engineering and Recombinant DNA Technology	DE	3	0	0	3
	ZOL.529	Genetic Engineering					
13	BCH.527	Developmental Biology	DE	3	0	0	3
	MME.555	Evolutionary and Developmental Biology					
Total				20	0	8	24

SEMESTER III							
S. No.	Course Code	Course Title	Course Type				
				L	T	P	Cr
1	CCC.551	Research Methodology	CC	3	0	0	3
2	BIM.524	Systems Biology	CC	2	0	2	3
3	BIM.525	Biological Databases and Management Systems	CC	3	0	0	3
4	BIM.526	Biological Databases and Management Systems Lab (Practical)	SBC	0	0	4	2
5	BIM.527	Biomolecular Simulations and Drug design Lab (Practical)	SBC	0	0	4	2
6	PCP.520	Entrepreneurship	CF	1	0	0	1
7	BIM.528	Enriching Bioinformatics	DEC	2	0	0	2
8	BIM.600	Dissertation Part-I	SBC	0	0	8	4
Total				11	0	18	20

SEMESTER IV							
S. No.	Course Code	Course Title	Course Type				
				L	T	P	Cr
1	BIM.601	Dissertation Part -II	SBC	0	0	40	20
Total				0	0	40	20
Grand Total				86 Credits			

Semester	L	T	P	Cr
I	19	0	6	22
II	20	0	8	24
III	11	0	18	20
IV	0	0	40	20
TOTAL	50	0	72	86

L: Lectures; T: Tutorial; P: Practical

MOOCs may be taken up to 40% of the total credits (excluding dissertation credits). MOOC may be taken in lieu of any course but content of that course should match a minimum 70%. Mapping will be done by the department and students will be informed accordingly.

Mode of Transaction: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Evaluation Criteria for Theory Courses

A. Continuous Assessment (Course wise): [25 Marks]

Two or more of the given methods (Surprise Tests, in-depth interview, unstructured interview, Jigsaw method, Think-Pair Share, Students Teams Achievement Division (STAD), Rubrics, portfolios, case based evaluation, video based evaluation, Kahoot, Padlet, Directed paraphrasing, Approximate analogies, one sentence summary, Pro and con grid, student generated questions, case analysis, simulated problem solving, media assisted evaluation, Application cards, Minute paper, open book techniques, classroom assignments, homework assignments, term paper).

B. Mid Semester Test: Based on Subjective Type Test [25 Marks]

C. End Semester Test: Based on Subjective (70%) and Objective (30%) Type Test [50 Marks]

The objective type will include one-word answers, fill-in the blank, sentence completion, true/false, MCQs', and matching, analogies. The subjective type will include a very short

answer (1-2 lines), short answer (one paragraph), essay type with restricted response, and essay type with extended response.

Core, Discipline Elective, Compulsory Foundation, Value Added and Interdisciplinary Courses			Discipline Enrichment Course		Entrepreneurship Course	
	Marks	Evaluation	Marks	Evaluation	Marks	Evaluation
Internal Assessment	25	Various	-	-	-	-
Mid-semester test (MST)	25	Subjective	50	Objective	25	Objective
End-semester test (EST)	50	Subjective (70%) Objective (30%)	50	Objective	25	Subjective

Evaluation Criteria for Practical Courses:

Evaluation	Marks
Maintaining the lab records/notebooks	10
Continuous assessment	20
Attendance	10
Final practical examination	50
Viva-voce	10

CF: Compulsory Foundation, CC: Core Course, DE: Discipline Elective, DEC: Discipline Enrichment Course, IDE: Inter-Disciplinary Elective, SBC: Skill-based Core, SBE: Skill-based Elective, VAC: Value Added Courses

L: Lecture, T: Tutorial, P: Practical

* Every student has to take 1 IDE (Inter-Disciplinary Elective) course of 2 credits from other disciplines in 1st semester of the program.

ZOL: Department of Zoology

LHG: Department of Human Genetics and Molecular Medicine

BOT: Dept of Botany

BCH: Dept. of Biochemistry

M.Sc. in Bioinformatics (Batch 2022)**Course Structure of the Programme****Total Credit: 86**

	Core courses	Elective courses			Foundation courses		Total Credit
		DE	ID	SB	CF	EF/VB	
Sem I	4 (12 Cr)	1 (3 Cr)	1 (2 Cr)	1 (2 Cr)	1 (3 Cr)	--	22
Sem II	3 (9 Cr)	2 (6 Cr)	--	2 (4 Cr)	1 (3 Cr)	1 (2 Cr)	24
Sem III	3 (9 Cr)	--	--	2 (4 Cr) 1 (4 Cr Dissertation)	1 (1 Cr Entre.) 1 (2 Cr DEC)	--	20
Sem IV	--	--	--	01 (20 Cr Dissertation)	--	--	20
Total Credit	30	9	2	34	9	2	86

DE: Discipline Elective, ID: Interdisciplinary, SB: Skill based (Practical); Dissertation, CF: Compulsory foundation, EF: Elective Foundation, VB: Value Based

SEMESTER-I

Course Title: Biological Sciences
Course Code: BIM.506
Course type: CC
Total Hours: 45

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to:

CLO1: Demonstrate the mechanism of various biomolecules to act in a cell or an organism

CLO2: Summarize the functional classification of proteins

CLO3: Discuss the structure and function of eukaryotic cell components and cell division

CLO4: Explain the DNA organization and differentiate the structures of DNA

CLO5: Describe the protein synthesis and gene expression process

CLO6: Describe structure, function and growth of microbes

CLO7: Apply the principles of rDNA technology to design gene cloning experiments in prokaryotes and eukaryotes

Units/ Hours	Contents	Mapping with CLO
I 15 Hours	<p>Biochemistry of biological macromolecules Carbohydrates-configurations, conformations, classification. Monosaccharides, disaccharides and polysaccharides- types, characteristics and properties. Central pathways of carbohydrate metabolism. Glycosaminoglycans, Glyco proteins-structure and function. Lipids-Classification, structure, properties; biological significance. Introduction to Lipid metabolism and Biosynthesis of fatty acids, lipid linked proteins, lipoproteins.</p> <p>Learning Activities: Peer discussion, seminars on recent studies on Carbohydrates and Lipids, visualisation and analysing the structures of Carbohydrates and Lipids, Peer discussion on the importance of the molecules.</p>	CLO1

	<p>Amino acids-classification and properties, isoelectric point. Peptides and proteins- functional classification of proteins-enzymes, transport, membrane, hormone, antibody, storage, structural etc. Amino Acids-biosynthetic pathways and their regulation. Enzyme classifications; Enzyme kinetics; Michaelis-Menten equation. Nucleic acid bases and Nucleotides, fundamentals of DNA and RNA.</p> <p>Learning Activities: Peer discussion, seminars on recent studies on proteins, Visualisation and analysing the protein structures, Peer discussion on the importance of protein functions.</p>	CLO2
II 12 Hours	<p>Cell and molecular biology Prokaryotes and eukaryotes, eukaryotic cell components and function, localization of genetic components - cytoskeleton in eukaryotes and their structure and composition – microtubules, microfilaments. Eukaryotic cell cycle - mechanics of cell division.</p> <p>Learning Activities: Peer discussion, group discussions, Research paper presentation, Class Quiz</p>	CLO3
	<p>DNA structure, DNA replication, Transcription machinery-RNA synthesis and processing, transcription factors, types of RNA and its structures, post-transcriptional processing of mRNA. ribosomes – molecular mechanism of translation machinery, translation in prokaryotes and eukaryotes – post translational modifications, Eukaryotic gene regulation.</p> <p>Learning Activities: Peer discussion on various DNA forms and its roles. Research paper presentation, Problem based learning sessions, Class quiz.</p>	CLO4 & CLO5
III 8 Hours	<p>Introduction to microbiology Importance of microorganisms, Bacterial cell structure and function, Microbial Metabolism, morphology and classification of major groups of microorganisms: fungi, bacteria, viruses, protozoa, algae, nematodes. Molecular taxonomy, Microbial Growth and Biofilms. Sterilization procedures, culture media preparation, types of culture media. Simple staining, negative staining, differential staining.</p> <p>Learning Activities: Demonstration at the cell culture facilities, Group discussion, research paper discussion</p>	CLO6
V 10 Hours	<p>Methods in Biology Molecular cloning of DNA or RNA fragments in bacterial and eukaryotic systems. PCR-principle and working concept, <i>in-silico</i> designing of specific and non-specific primers, validation of primers design, troubleshooting with primers.</p>	CLO7

	<p>construction of mutants, protein overexpression and purification. RFLP, CRISPR-CAS system for genome engineering.</p> <p>Learning Activities: Visit the molecular biology labs to and demonstrate various aspects on molecular cloning, protein purification, Group discussion, research paper discussion, Class quiz</p>	
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Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Berg, J.M., Tymoczko, J.L. and Stryer, L. (2019). Biochemistry 9th edition. W.H. Freeman & Company. USA, ISBN: 9781319248062
2. Nelson, D. and Cox, M.M. (2021). Lehninger Principles of Biochemistry. W H Freeman & Co; 8th edition, ISBN-10: 1319228003
3. Hoerter, J. E., & Ellis, S. R. (2020). Biochemistry, Protein Synthesis. In StatPearls. StatPearls Publishing.
4. Voet, D. and Voet, J.G. (2014). Principles of biochemistry. CBS Publishers & Distributors. New Delhi, India, ISBN-10: 0470570954
5. Venki Ramakrishnan, Jennifer A Doudna (2019). Gene Machine: The Race to Decipher the Secrets of the Ribosome. ISBN: 9781786076717. Oneworld Publications
6. Mercadante, A. A., Dimri, M., & Mohiuddin, S. S. (2020). Biochemistry, Replication and Transcription. In StatPearls. StatPearls Publishing.

Course Title: Sequence Analysis
Course Code: BIM.507
Course type: CC
Total Hours: 45

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to:

CLO1: Apply the algorithms for pairwise and multiple alignment

CLO2: Choose suitable scoring matrix for sequence alignment

CLO3: Perform multiple sequence alignment

CLO4: Perform searching in sequence database and identify the motifs and profiles in the given biological sequences

Units/ Hours	Contents	Mapping with CLO
I 13 Hours	Basic concepts of sequence similarity, identity and homology, homologues, orthologues, paralogues and xenologues Pairwise sequence alignments: basic concepts of sequence alignment, Needleman and Wunsch, Smith and Waterman algorithms for pairwise alignments, gap penalties. Learning Activities: Peer discussion, hands-on training in global and local alignments, Problem based learning.	CLO1
II 10 Hours	Scoring matrices: basic concept of a scoring matrix, PAM and BLOSUM series, Comparison of PAM and BLOSUM, Tools such as BLAST (various versions of it) and FASTA. Learning Activities: Hands-on training in PAM and BLOSUM matrix, Classroom presentation, Class quiz.	CLO2
III 12 Hours	Multiple sequence alignments (MSA): basic concepts of various approaches for MSA (e.g., progressive, hierarchical etc.). Algorithm of CLUSTALW (including interpretation of results), concept of dendrogram and its interpretation. Learning Activities: Research paper presentation, Peer discussion on various algorithms of MSA.	CLO3
IV 10 Hours	Sequence patterns and profiles: Basic concept and definition of sequence patterns, motifs and profiles, profile-based database searches using PSI- BLAST, analysis and interpretation of profile-based searches. troubleshooting with primers. construction of mutants, protein overexpression and purification. RFLP, CRISPR-CAS system for genome engineering. Learning Activities: Case studies, research paper discussion, Student-generated test questions.	CLO4

Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. David W. Mount (2001). Bioinformatics. Cold Spring Harbor Laboratory Press, ISBN 0-87969-608-7
2. Pevzner P. A. (2004). Computational Molecular Biology. Prentice Hall of India Ltd, ISBN 81-203-2550-8
3. Krane D.E. and Raymer M.L. (2003). Fundamental concepts of Bioinformatics. Pearson Education ISBN 81-297-0044-1
4. Gautham N. (2006). Bioinformatics. Narosa publications ISBN-13: 9781842653005
5. Gayathri N. Silva (2021). Bench guide for Protein expression and Purification. Bluerose Publishers Pvt. Ltd, ISBN-10: 935427725X.
6. Labrou, N. E., and Labrou. (2021). Protein downstream processing. Springer US. ISBN: 978-1-62703-977-2.
7. Singh, V., and Dhar, P. K. (2020). Genome engineering via CRISPR-Cas9 system. Academic Press. ISBN: 9780128181409.
8. Baxevanis, A. D., Bader, G. D., & Wishart, D. S. (2020). Bioinformatics: A Practical Guide to the Analysis of Genes and Proteins 4th Edition. Wiley Publishers, ISBN-10: 1119335582
9. Yoon B.J., Qian, X (2021). Recent Advances in Biological Network Analysis. Springer International Publishing, ISBN 978-3-030-57172-6.

Course Title: Structural Bioinformatics
Course Code: BIM.508
Course type: CF
Total Hours: 45

L	T	P	Cr
2	0	2	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to:

CLO1: Describe the macromolecular structures and the experimental methods to determine it

CLO2: Apply various structure analysis tools to assign protein fold, biological function and analyse the intramolecular interactions.

CLO3: Predict the protein tertiary structures from its amino acid sequence.

CLO4: Identify the membrane bound regions, stabilizing interactions and biological function using computational tools

CLO5: Analyse various forces, which stabilize macromolecular structure and biological assemblies.

CLO6: Describe the role of structural genomics in explaining life process and drug discovery

Units/ Hours	Contents	Mapping with CLO
I 12 Hours	<p>Macromolecular Structures Introduction to biological macromolecules, Structural organization of proteins, Forces Stabilizing Proteins, Structure determination methods - X-ray crystallography, NMR, Cryo-electron microscopy, small angle X-ray scattering and Neutron scattering. Introduction to PDB Data-file formats, visualizing Structures, reading coordinate files, potential challenges. Structure validation - Ramachandran plot. Software for quality check.</p> <p>Learning Activities: Peer discussion, visualization and analysing of protein structures, training on various structural databases and validation, Visit and demonstration of X-ray, NMR and EM facilities.</p>	CLO1
II 10 Hours	<p>Structure comparison and alignment Impact of protein structure comparison and alignment, sequence-structure relationship, general approaches, statistical analysis Structure comparison, multiple structure alignment, software/tools for structure comparison and alignment</p> <p>Structure-function relationship Secondary structure assignment methods, Structural classification of proteins, Structural domains, Relationship between structure and function, assigning function from structure.</p>	CLO2

	<p>Learning Activities: Problem based learning, Students Teams, hands-on training on structure comparison and derive structure-function relationships, Case studies, research paper discussion</p>	
III 12 Hours	<p>Secondary structure prediction methods, Protein tertiary structure prediction- Comparative modeling, Threading, <i>Ab initio</i> modeling, introduction to artificial intelligence, Deep-learning algorithms in protein structure prediction, CASP experiments.</p> <p>Learning Activities:, Hands-on training structure prediction, Peer group discussion, Student seminars on recent developments</p>	CLO3
	<p>Membrane Protein Structure Prediction, Transmembrane region prediction, difficulty in solving membrane protein structures; structural genomics of membrane proteins; tools and databases for identification of membrane proteins and the prediction of their structures.</p> <p>Learning Activities: Hands-on training on membrane structure prediction, Peer group discussion on structural genomics findings, Student seminars on recent developments</p>	CLO4
IV 11 Hours	<p>Protein-Protein Interaction, evolutionary features related to structure and function, prediction of interacting regions and interaction partners. Structural analysis on protein-DNA interactions, protein-DNA binding specificity, Inter- and intramolecular interactions in protein-DNA recognition-prediction of DNA-binding sites, databases and tools for protein-DNA interactions. Introduction to RNA Structural Bioinformatics</p> <p>Learning Activities: Peer discussion, Visualizing Protein-protein/DNA interactions, Research paper presentation, Problem based learning.</p>	CLO5
	<p>Structural Genomics Structural annotation of genomes and structural genomics, structural genomics initiatives, impact of structural genomics on drug discovery, structural genome annotation resources.</p> <p>Lear Learning Activities: Peer discussion, Exploring the outcomes of various Structural genomics initiatives, Case studies on the role of structural genomics in drug discovery.</p>	CLO6

Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Branden, C. and Tooze, J. (1999). Introduction to Protein Structure, Garland Publishing Inc., 2e
2. Pal, S. (2020). Fundamentals of Molecular Structural Biology. Academic Press. ISBN: 9780128148556.
3. Gu, J. and Bourne, P. E. (2009) Structural Bioinformatics, John Wiley & Sons, 2e,
4. Liljas, A and L. Liljas, J. Piskur, G. Lindblom, P. Nissen and M. Kjeldgaard. (2009) Textbook of Structural Biology. World Scientific Publishing Co.
5. Rupp, B. (2009) Biomolecular Crystallography: Principles, Practice, and Application to Structural Biology. Garland Science.
6. Gromiha, M.M., (2010) Protein Bioinformatics, From Sequence to Function, Academic Press, USA, 1e.
7. Zhou, Y., Kloczkowski, A., Faraggi, E., and Yang, Y. (2017). Prediction of protein secondary structure. Humana Press. ISBN: 9781493964048, 9781493964048.
8. Kihara, D. (2014). Protein structure prediction. New York, NY, USA: Humana Press. ISSN: 1064-3745.
9. Anderson, W. F. (2014). Structural genomics and drug discovery. Methods and Protocols, 1. Humana Press. ISBN: 9781493903535, 9781493903535.

Web resources:

NPTL <http://nptel.ac.in/syllabus/syllabus.php?subjectId=104102016>

MIT OpenCourseWare

<http://ocw.mit.edu/courses/biological-engineering/20-442-molecular-structure-of-biological-materials-be-442-fall-2005/>

Course Title: Python Programming

Course Code: PCP.519

Course type: CC

Total Hours: 45

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to:

CLO1: Describe Python Programming to build applications in their core domain.

CLO2: Write python scripts by using decision making command and loops

CLO3: Apply functions, modules and external packages in Programming

CLO4: Write programs by calling inputs and write specific output files

Units/ Hours	Contents	Mapping with CLO
I 12 Hours	Introduction, Data Types and Operators: Installation and working with Python, Variables and data types in python, perform computations and create logical statements using Python's operators: Arithmetic, Assignment, Comparison, Logical, Membership, Identity, Bitwise operators, list, tuple and string operations Learning Activities: Peer discussion, brain storming and Problem based learning sessions, Class quiz	CLO1
II 11 Hours	Python Decision making and Loops: Write conditional statements using If statement, if ...else statement, elif statement and Boolean expressions, while loop, for loop, Nested Loop, Infinite loop, Break statement, continue statement, Pass statement, Use for and while loops along with useful built-in functions to iterate over and manipulate lists, sets, and dictionaries. Plotting data, Programs using decision making and loops Learning Activities: Peer discussion on different code designing, Decoding quiz	CLO2
III 11 Hours	Python Functions and Modules: Defining custom functions, Organizing Python codes using functions, Create and reference variables using the appropriate scope, Basic skills for working with lists, tuples, work with dates and times, get started with dictionaries, importing own module as well as external modules, Programming using functions, modules and external packages Learning Activities: Brainstorming and Problem Solving code designing.	CLO3
IV 11 Hours	Python File Operations: An introduction to file I/O, use text files, use CSV files, use binary files, handle a single	CLO4

	exception, handle multiple exceptions, Illustrative programs. Learning Activities: Problem Solving code practice based on CSV files.	
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Transaction Mode: Lecture, tutorial, problem solving

Suggested Readings

1. Gowri Shankar, S., Veena, A. (2018). Introduction to Python Programming, 1st Edition, CRC Press/Taylor & Francis.
2. VanderPlas, J. (2016). Python Data Science Handbook: Essential Tools for Working with Data, 1st Edition, O'Reilly Media.
3. Géron, A. (2019). Hands-On Machine Learning with Scikit-Learn and TensorFlow: Concepts, Tools, and Techniques to Build Intelligent Systems, 2nd Edition, O'Reilly Media.
4. Chun, W. J. (2015). Core Python Applications Programming, 3rd Edition, Pearson Education India.

Course Title: Genomics and Proteomics
Course Code: BIM.509
Course type: CC
Total Hours: 45

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to:

CLO1: Describe the DNA-Protein Interactions and Transcription process

CLO2: Use bioinformatics-OMIM database and identify SNPs

CLO3: Describe integrated genomic maps, gene expression profiling

CLO4: Apply probabilistic modeling techniques for building transcriptional regulatory networks

Unit/ Hours	Contents	Mapping with CLO
I 12 Hours	DNA- Protein interaction: Process of DNA-Protein Interactions During Transcription, Network identification and processing. Protein- Protein Interactions: Processing of the Proteome, Prediction of Post-translational modifications, Protein Degradation. Learning Activities: Classroom presentation and discussion, hands-on training on protein-DNA interaction analysis, Problem based learning.	CLO1
II 10 Hours	Role of bioinformatics-OMIM database, integrated genomic maps, gene expression profiling; identification of SNPs, Vector Screening. Learning Activities: Research paper discussion, Hands-on training on biological databases, Problem based learning.	CLO2
III 12 Hours	DNA microarray: database and basic tools, Gene Expression Omnibus (GEO), understanding of microarray data, normalizing microarray data, Raw read quality assessment, Mapping data visualization, detecting differential gene expression. Learning Activities: Peer discussion, Hands-on training on Genomic databases and data visualization, Problem based learning.	CLO3
IV 11 Hours	Building predictive models of transcriptional regulatory networks using probabilistic modeling techniques. Viral Informatics: Virome, Comparative metagenomic analysis using VIROME. Metagenomics	CLO4

	<p>Learning Activities: Peer discussion on probabilistic approaches for transcriptional networks, Hands-on training on biological networks, quiz.</p>	
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Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Suhai, S (2002). Genomics and Proteomics. Springer US
2. Campbell (2007). Discovering Genomics, Proteomics and Bioinformatics. Pearson Education
3. Informatics, G., and Berger, S. (2021). Viral Meningitis: Global Status: 2021 edition. GIDEON Informatics Inc. ISBN: 1498833098, 9781498833097.
4. Lesk, A. (2019). Introduction to bioinformatics. Oxford university press. ISBN: 9780198794141.
5. Grant, P.R. (2004). Computational Genomics: Theory and Application. Horizon Bioscience
6. Gracia ,K.C, Husi H. (2019) Computational Approaches in Proteomics. Brisbane (AU): Codon Publications, PMID: 31815395.
7. Guzzi, P.H. (2016). Microarray Data Analysis: Methods and Applications. Humana Press. ISBN: 1493931725, 9781493931729.

Web Resources

1. <http://virome.dbi.udel.edu>

Course Title: Genomics and Proteomics Lab (Practical)
Course Code: BIM.510
Course type: SBC
Total Hours: 60

L	T	P	Cr
0	0	4	2

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Perform database search and retrieve genomic and proteomic data for analysis

CLO2: Utilize the tools available to analyse genome and protein sequences

CLO3: Identify different types of protein–DNA interactions and characterize different biological networks.

CLO4: Perform protein structure classification

CLO5: Perform protein structure prediction and function from the amino acid sequence

Course Content

The following experiments should be conducted by the students:

A. Genomics

1. Retrieve the gene sequences by exploring and querying the nucleic acid databases.
2. Find the chromosomal location of gene sequence and basic experiments in NCBI map viewer.
3. Genome Annotation i.e. Mining Genomic Sequence Data, gene prediction methods
4. Gene expression data analysis :Differential gene expression analysis, Peak finding, Motif Discovery.

B. Proteomics

1. Retrieve the protein sequences by exploring and querying the protein databases.
2. Select a protein family to find out its superfamily. Find out structural and functional information about above protein families and superfamily
3. Use on-line motif identification tools to predict motif in a set of sequences
4. Secondary structure Prediction methods and comparisons of various secondary structure prediction tools
5. Prediction of Protein- protein interaction

Transactional Modes: Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Mani, K. & Vijayaraj-Aparna, N. Bioinformatics: A Practical Approach- Publishers New Delhi
2. Informatics, G., and Berger, S. (2021). Viral Meningitis: Global Status: 2021 edition. GIDEON Informatics Inc.
3. Lesk, A. (2019). Introduction to bioinformatics. Oxford university press.
4. Deitel, P. J. (2002). How to program, Pearson

Course Title: Protein Engineering
Course Code: BIM.511
Course type: DE
Total Hours: 45

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to:

CLO1: Describe the protein folding process, mechanism and its importance.

CLO2: Choose a suitable experimental technique to introduce desired amino acid modifications to wild type proteins

CLO3: Design and modify sequence for a protein with a desired structure and/or property

CLO4: Design the protein molecules with the desired bonded and non-bonded interactions using computational methods

Units/ Hours	Contents	Mapping with CLO
I 10 Hours	<p>Protein Folding and stability: Protein structural features, protein structure-function relationship, Protein Folding: Theory and Experiment-Protein Renaturation, Determinants of Protein Folding, Folding Pathways, Folding Accessory Proteins. Introduction to Conformational Diseases. Protein stabilising factors, Protein Denaturation, Explaining the Stability of Thermostable Proteins</p> <p>Learning Activities: Peer discussion, demonstration using physical and computer models on folding process, Student seminars</p>	CLO1
II 13 Hours	<p>Strategies for Protein Design: Introduction to protein expression and mutagenesis. Protein engineering using unnatural amino acids- methodologies; applications-enhanced stability, tuning catalytic activity, tuning selectivity, enzyme design</p> <p>Protein design; strategies for the design of structure - self-assembly - ligand-induced assembly - assembly via covalent cross-linking - assembly of peptides on a synthetic template. Strategies for the design of function-novel functions by retrofitting natural proteins - incorporation of binding sites into <i>de novo</i> proteins - design of catalytically active proteins - membrane proteins and ion channels - design of new materials.</p> <p>Learning Activities: Peer discussion, Case studies, research paper discussion, Problem based learning</p>	CLO2

III 10 Hours	<p>Computational Protein Design: Methods of Computational Protein Design, core and full repacking, predicting native protein core sequences; altering protein folds. Geometry and stereochemistry-based design, Case studies on Computationally Designed Proteins.</p> <p>Learning Activities: Problem based learning, demonstration using and computer models and experimental evidences, Classroom presentation</p>	CLO3
IV 12 Hours	<p>Engineering artificial metalloenzymes, engineered cytochromes P450 for Biocatalysis, Application of engineered biocatalysts for the synthesis of active pharmaceutical ingredients, Engineering antibody-based therapeutics: progress and opportunities, Development of novel cellular imaging tools using protein engineering.</p> <p>Learning Activities: Peer discussion, Case studies, research paper discussion, Problem based learning</p>	CLO4

Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Zhao, H. (2021) Protein Engineering: Tools and Applications, Wiley-VCH Verlag GmbH & Co
2. Park, S.J., and Cochran, J.R. (2010). Protein Engineering and Design, 1/e, Taylor and Francis Inc., CRC Press, USA.
3. Carey, P.R. (1996) Protein Engineering and Design, 1/e, Academic Press Inc, USA.
4. Samish, I. (2017). Computational Protein Design, 1/e, Humana Press, New York.
5. Branden, C. I., & Tooze, J. (1999)., Introduction to Protein Structure, 2/e, Garland Science, USA.
6. Stefan, L. and Uwe, T.B. (Eds), (2012) Protein Engineering Handbook: Volume 3, 1/e, Wiley-VCH Verlag GmbH & Co.

Web resources:

NPTEL

1. https://onlinecourses.nptel.ac.in/noc21_cs100/preview
2. https://onlinecourses.nptel.ac.in/noc21_bt14/preview

MIT Open Courseware

1. <https://ocw.mit.edu/courses/biology/7-344-antibiotics-toxins-and-protein-engineering-spring-2007/>
2. <https://ocw.mit.edu/courses/biology/7-91j-foundations-of-computational-and-systems-biology-spring-2014/video-lectures/lecture-12-introduction-to-protein-structure-structure-comparison-and-classification/>

Course Title: Linux and Perl
Course Code: BIM.529
Course type: DE
Total Hours: 45

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Demonstrate Linux packages and use various compressed file formats

CLO2: Describe Perl programming to build application

CLO3: Demonstrate web-based PERL CGI programming concepts

CLO4: Develop scripts for Bioinformatics applications using PERL programming.

Units/ Hours	Contents	Mapping with CLO
I 15 Hours	<p>Basics of Linux shell, BASH, CSH, PERL Basics Scalar variables, Syntax and semantics, Processing scalar variables, Iteration with while construct, Variable containers, Loops, Conditional statements, Introducing Patterns, Reading and writing files, Case study: Making Motif Search tool.</p> <p>Learning Activities: Peer discussion, brain storming and Problem based learning sessions, Class quiz</p>	CLO1
II 11 Hours	<p>Advance data structure and programming in PERL Arrays, Hashes, Sub-routines, Regular expression and Text mining: The Match Operator, Match Operator Modifiers, The Substitution Operator, Substitution Operator Modifiers, Translation, Translation Operator Modifiers, More complex regular expressions</p> <p>Learning Activities: Peer discussion on different code designing, Decoding quiz</p>	CLO2
III 8 Hours	<p>PERL CGI Programming Introduction to HTTP, HTTP Methods, generating web pages with Perl, Generating web pages with Perl, CGI. pm, HTML forms and input fields.</p> <p>Learning Activities: Brainstorming and Problem Solving code designing.</p>	CLO3
IV 10 Hours	<p>Bioperl Introduction to Bioperl, creating a sequence and an object, manipulating sequence data with Seq methods, writing a sequence to a file, retrieving a sequence from a file, retrieving a sequence and multiple sequences from a database. Obtaining basic sequence statistics, Identifying restriction enzyme sites, Identifying amino acid cleavage sites, Running applications: BLAST, Searching for genes in genomic DNA, Using EMBOSS applications with Bioperl, Aligning sequences with Smith-Waterman.</p> <p>Learning Activities: Peer discussion on different code designing, Decoding quiz</p>	CLO4

Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning.

Suggested Reading

1. Moorhouse M, Barry P (2005): Bioinformatics Biocomputing and Perl: An Introduction to Bioinformatics Computing
2. Skills and Practice, Book, John Wiley & Sons
3. Dwyer R. A. (2003): Genomic Perl: From Bioinformatics Basics to Working Code, Volume 1, Book, Cambridge University Press
4. Tisdall J (2003): Mastering Perl for Bioinformatics, Book, O'Reilly
5. Hietaniemi J, John Macdonald J, Orwant J (1999): Mastering Algorithms with Perl, Book, O'Reilly
6. William "Bo" Rothwell (2020) Advanced Perl Programming: From Advanced to Expert, Apress, 1st ed.
7. John Bach (2020) Programming Perl: The Ultimate Beginner's Guide to Learn Perl Programming Step by Step.
8. Rosenfeld L, Downey A.B, (2017) Think Perl 6: How to Think Like a Computer Scientist, O'Reilly Media

Web Resources

1. Bradnam K & Korf I (2012): Unix and Perl Primer for Biologists, Web tutorial at http://korflab.ucdavis.edu/Unix_and_Perl/current.html
Robert's PERL tutorial <http://www.physics.rutgers.edu/~kotliar/>

Course Title: Introduction to Structural Biology**Course Code: BIM.513****Course type: IDC****Total Hours: 30**

L	T	P	Cr
2	0	0	2

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Describe the physical and structural properties of proteins and the forces that stabilize protein structure

CLO2: Describe the principles and the practical aspects of macromolecular crystallography

CLO3: Describe the principles of Macromolecular NMR spectroscopy and Cryo EM and their application to study the structures of macro molecules

CLO4: Describe the applications bioinformatics tools in macromolecular structure determination.

Units/ Hours	Contents	Mapping with CLO
I 8 Hours	<p>Introduction to Protein structure: Protein structure: Amino acids and their properties, peptide bond, primary, secondary and tertiary structure - structural properties required for complex formation, Ramachandran Plot, Stabilizing forces, evolutionary conservation of structure-function in proteins.</p> <p>Learning Activities: Demonstration using physical and computer models, Student seminars, Peer group discussion</p>	CLO1
II 8 Hours	<p>MACROMOLECULAR CRYSTALLOGRAPHY Protein crystallization, Introduction to X-ray sources and detectors, X-ray diffraction - Bragg's law, structure factor and electron density map, phase problem, Molecular Replacement, experimental phasing techniques, structure validation.</p> <p>Learning Activities: Peer discussion, Visit and demonstration of X-ray diffractometer, demonstration using protein models, Student seminars</p>	CLO2
III 7 Hours	<p>NMR SPECTROSCOPY Principles of NMR spectroscopy, Chemical shift, Relaxation parameters, spin-spin coupling; multidimensional NMR; NMR Structure determination.</p> <p>CRYO ELECTRON MICROSCOPY Basic scanning and transmission microscopy, Introduction to cryo EM, determining the structure using cryo EM.</p>	CLO3

	Learning Activities: Peer discussion, Visit and demonstration of NMR and EM facilities, Classroom presentation, Peer group discussion	
IV 7 Hours	<p>STRUCTURAL BIOINFORMATICS Homology modelling, Fold recognition methods, <i>ab initio</i> methods, Prediction of membrane helices and accessibility. Introduction to structural genomics</p> <p>Learning Activities: Hands-on training on the bioinformatics tools, Peer discussion, research paper discussion</p>	CLO4

Suggested Reading

1. Kessel, Amit and Nir Ben Tal, *Introduction to Proteins: Structure, Function and Motion*, CRC Press, 2e, 2018.
2. Pal, Subrata, *Fundamentals of Molecular Structural Biology*, Academic Press, 2020.
3. Anders Liljas, Lars Liljas , Jure Piskur, Göran Lindblom, Poul Nissen, Morten Kjeldgaard, *Textbook of Structural Biology*, World Scientific Publishing Co Pte Ltd, 2/e, 2017
4. Timir Tripathi and Vikash Kumar Dubey, *Advances in Protein Molecular and Structural Biology Methods*, Academic Press, 2022

Web Resources

1. <https://ocw.mit.edu/courses/biology/7-91j-foundations-of-computational-and-systems-biology-spring-2014/video-lectures/lecture-12-introduction-to-protein-structure-structure-comparison-and-classification/>
2. <https://nptel.ac.in/courses/102107086>

SEMESTER II

Course Title: Biomolecular Simulations and Drug Design

Course Code: BIM.514

Course type: CC

Total Hours: 45

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Describe the modelling of small to large molecular environments

CLO2: Describe the concept of molecular mechanics force fields and select an appropriate energy function/force field for a given problem

CLO3: Choose an appropriate energy minimization method for the required simulation study.

CLO4: Apply different methods for simulating large molecular systems

CLO5: Describe a pharmacophore model from a set of drug molecule and quantify the structure activity relationship

CLO6: Perform and evaluate different virtual screening methods using large datasets

Units/ Hours	Contents	Mapping with CLO
I 11 Hours	<p>Molecular Modeling and Structure Introduction to Molecular modelling, Coordinate systems, potential energy surfaces for simple molecules. mini tutorials with protein and nucleic acid structure as examples.</p> <p>The simple molecular mechanics force field; general features of molecular mechanics force fields. Force Fields and Molecular Representation – Intramolecular Interactions, Non-bonded Interactions – London (van der Waals) Interactions, Electrostatic Interactions, Hydrogen Bonds, Constraints and Restraints, United Atom and Other Coarse-Grained Approaches, Non-pairwise Interactions, accuracy of the force fields.</p> <p>Learning Activities: Peer discussion, Demonstration using physical and computer model, Problem based learning, Analysing Experimental data.</p>	CLO1 & CLO2
II 12 Hours	<p>Energy Minimization and Related Analysis Techniques Steepest Descent, Conjugate Gradient, Newton-Raphson, Comparison of Methods, Advanced Techniques: Simulated Annealing, Branch-and-bound, Simplex, What's the big deal about the minimum?</p> <p>Phase space, Ergodicity, and Liouville's theorem, Ensemble theory, Thermodynamic averages -</p>	CLO3

	<p>Microcanonical Ensemble, Canonical Ensemble, Other MD Simulation Related Ensembles</p> <p>Learning Activities: Hands-on training, Peer discussion, Research paper presentation</p>	
<p>III 11 Hours</p>	<p>Methods for Simulating Large Systems Non-bonded Cutoffs – Shifted Potential and Shifted Force, Switching Functions, Neighbor Lists. Boundaries – Periodic Boundary Conditions, Stochastic Forces at Spherical Boundary. Long-range Interactions – The Ewald Sum, The Reaction Field Method</p> <p>Molecular dynamics using simple models, Molecular dynamics with continuous potentials, finite difference and predictor-corrector integration methods, choosing the time step. Calculating properties by integration; implementation of the Metropolis Monte Carlo method; Monte Carlo simulation of rigid and flexible molecules; Lattice and continuous polymer models.</p> <p>Learning Activities: Hands-on training on simulation, Peer discussion, Case studies, research paper discussion.</p>	CLO4
<p>IV 11 Hours</p>	<p>Drug design Introduction to drug designing, ADMET, drug metabolism, toxicity and pharmacokinetics. Lipinski rule of 5, Identification and validation strategies. Drug Target classification, Concept of Pharmacophore, Functional group considered as pharmacophore, Structure-based drug design, docking, QSAR, Artificial intelligence in drug discovery and development</p> <p>Learning Activities: Peer discussion, Case studies, research papers, Hands-on training on drug designing</p>	CLO5 & CLO6

Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Leach R. (2001). Molecular Modelling Principles and Applications 2nd Edition. Pearson.
2. Frenkel D. and Smit B. (2001). Understanding Molecular Simulation 2nd Edition, Academic Press.
3. Buschmann, H., & Holenz, J. (2018). Biomolecular Simulations in Structure-Based Drug Discovery. Germany: Wiley. ISBN:9783527342655
4. Alavi S. (2020). Molecular Simulations: Fundamentals and Practice 1st Edition, Wiley-VCH.

5. Bhatt, T. K., & Nimesh, S. (Eds.). (2021). The design and development of novel drugs and vaccines: Principles and protocols. Academic Press. ISBN: 9780128214718.
6. Renaud, J. P. (Ed.). (2020). Structural biology in drug discovery: Methods, Techniques, and Practices. John Wiley & Sons. ISBN: 9781118900406.
7. Allen, M. P., & Tildesley, D. J. (2017). Computer Simulation of Liquids 2nd Edition, Oxford University Press.
8. Stromgaard, K., Krogsgaard-Larsen, P., & Madsen, U. (2016). Textbook of Drug Design and Discovery. CRC Press, 5e, 2016.

Course Title: Data Mining and Machine Learning
Course Code: BIM.515
Course type: CC
Total Hours: 45

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Perform data cleaning, cross-validation, and application of regression analysis

CLO2: Use various methods of clustering and dimensionality reduction for data analysis

CLO3: Explain classification as a tool to develop predictive platform

CLO4: Apply SVM and Neural network methods data analysis.

Units/ Hours	Contents	Mapping with CLO
I 10 Hours	<p>Introduction: Overview of Machine Learning field, Artificial intelligence Vs Machine learning, Regression and classification problem, cocktail party problem and algorithm, cross validation, terminology alert: true positive, false positive, Specificity, Sensitivity, Accuracy, Recall, Precision, AUC-ROC curves, Confusion matrix, Errors in ML: Bias and variance.</p> <p>Learning Activities: Demonstration using random dataset, Problem based learning,</p>	CLO1
II 12 Hours	<p>Unsupervised Methods: Clustering: Distance Metrics, K-Means, leader, Jarvis-Patrick, hierarchical clustering; Dimensionality Reduction: Principal Component Analysis (PCA), Linear Discriminant Analysis (LDA), Partial Least Squares – Discriminant Analysis (PLS-DA).</p> <p>Learning Activities: Peer discussion, Problem based learning, Analysing Experimental data for unsupervised model training.</p>	CLO2
III 12 Hours	<p>Supervised Methods: Univariate and multivariate linear regression: Model representation and cost function; Classification: k-nearest neighbors' algorithm (K-NN), naïve Bayes, decision trees, boosting and bagging, Bonferroni Correction.</p> <p>Learning Activities: Case study, Peer discussion, brainstorming and Problem Solving.</p>	CLO3
IV 11 Hours	<p>Other ML algorithms: Ensemble methods, random Forests; Support vector machines Neural networks, Recommendation systems; Outlier detection.</p>	CLO4

	<p>Learning Activities: Problem Solving, case study, Peer discussion on various classification methods, brainstorming discussion</p>	
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Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Helder I., N. (2021). Bioinformatics. Exon Publications.
2. Alonso-Betanzos, A., & Bolón-Canedo, V. (2018). Big-Data Analysis, Cluster Analysis, and Machine-Learning Approaches. *Advances in experimental medicine and biology*, 1065, 607–626.
3. Applied Predictive Modeling by Max Kuhn and Kjell Johnson; 2013.
4. James, G., Witten, D., Hastie, T., & Tibshirani, R. (2014). An introduction to statistical learning: With applications in R.
5. McKinney, W. (2013). Python for data analysis
6. Han, J., Kamber, M., & Pei, J. (2011). *Data Mining: Concepts and Techniques*, Third Edition.

Course Title: NGS sequence data analysis
Course Code: BIM.516
Course type: CF
Total Hours: 45

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Describe the experimental procedures involved in DNA sequencing

CLO2: Compare various NGS chemistries and NextGen Sequencing platforms

CLO3: Explain the principles and application of various sequencing methods.

CLO4: Describe the different workflows in the analysis of NGS data

CLO5: Describe complexity of NGS data, steps involved in NGS data analysis, and algorithms used for this purpose

Units/ Hours	Contents	Mapping with CLO
I 11 Hours	<p>Next-Generation Sequencing NGS basic terminology: genome, transcriptome, exome, metabolome and metagenome; evolution of sequencing technologies: first, second and third generation; sequencing methods; Samples and preparation; different NGS platforms (Roche 454, AB SOLiD, Illumina, Ion Torrent, PacBio, Nanopore sequencing); challenges in the NGS field; sanger vs NGS sequencing; read length and error rates.</p> <p>Learning Activities: Peer discussion, Seminars on application of the techniques, Visit and demonstration of NGS platforms</p>	CLO1 & CLO2
II 12 Hours	<p>NGS methods Genomics: Whole genome sequencing, Target sequencing, Exome sequencing, Pooled Sequencing and De novo sequencing; Transcriptomics: Total RNA and mRNA seq, targeted RNA-seq and small noncoding RNA-seq, single-cell RNA-sequencing (scRNA-seq); Epigenomics: Methylation sequencing (Methylated DNA immunoprecipitation sequencing (MeDIP-Seq), bisulfite sequencing (BS-seq), ChIP-Seq.</p> <p>Learning Activities: Discussion and seminars on application of the techniques, Case studies, research paper discussion</p>	CLO3
III 11 Hours	<p>NGS data Analysis workflow and file formats Primary, secondary and Tertiary analysis; Sequence quality evaluation - Phred Scores; Quality Control and Preprocessing; post alignment quality and bias control;</p>	CLO4

	Types of NGS data; File formats - FASTQ, SAM, BAM, VCF; Online NGS databases. Learning Activities: Hands-on training in sequence analysis, Seminars on application of the techniques, Case studies, research paper discussion	
IV 11 Hours	NGS Data Analysis RNA-Seq data analysis-expression estimation, Differential Expression Analysis Workflow; Aligning Reads to Reference; Transcriptome Assembly; Alignment programs; Spliced alignment Quantitation and Annotation-Based Quality Control, data visualization, Downstream analysis such as pathway analysis, clustering and Gene ontology. Learning Activities: Peer discussion, hands-on training in sequence analysis, Seminars on application of the techniques, Case studies, research paper discussion	CLO5

Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings:

1. Brown, S. M. (2013). Next-Generation DNA Sequencing Informatics. Cold Spring Harbor Laboratory Press, 1/e.
2. Xinkun Wang. (2016). Next-Generation Sequencing Data Analysis. Talyor and Francis Group.
3. Low, L., & Tammi, M. (2017). Introduction to next generation sequencing technologies. Bioinformatics: A Practical Handbook Of Next Generation Sequencing And Its Applications.
4. Korpelainen et al (2015). RNA-seq Data Analysis – A Practical Approach, CRC Press, Taylor & Francis Group, 1e.
5. Jianping Xu. (2014). Next-generation sequencing: Current Technologies and Applications. Caister Academic Press, 1/e.

Course Title: Biostatistics using R
Course Code: BIM.517
Course type: CC
Total Hours: 45

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Use the measures of central tendency dispersion on a given data and determine different statistical measures for the data

CLO2: Use correlation and linear regression methods to find a relationship and good of fit for the given data

CLO3: Perform hypothesis testing on small and large data samples using appropriate parametric and non-parametric test.

CLO4: Perform statistical hypothesis testing by selecting an appropriate testing procedure for the given analysis.

Estimate confidence intervals for a statistical parameter

CLO5: Calculate probabilities and estimate parameters for various outcomes/events following different probability distributions

Units/ Hours	Contents	Mapping with CLO
I 11 Hours	<p>Statistical Measures Types of biological data: Accuracy and Significant figures - Frequency distributions - Cumulative frequency distributions - Measures of central tendency - Mean, Median and Mode - Measures of dispersion- Range, Quartile deviation, Mean deviation and Standard deviation - Coefficient of variation- Skewness and Kurtosis. Linear correlation, Simple Regression.</p> <p>Learning Activities: Peer discussion, Case studies, research paper discussion</p>	CLO1 & CLO2
II 12 Hours	<p>Sampling Theory and Testing of Hypothesis Population - Sample - Standard Error - Sampling Distributions- Central limit theorem - Confidence intervals. Hypothesis testing: Null Hypothesis - Alternate Hypothesis - Single and Two tailed Hypothesis - Type I and Type II errors -The p value - A single population mean - The difference between two population means - Paired comparisons - A single population proportion - The difference between two population proportions - Testing the independence of attributes by chi-square test - Nonparametric test - Mann-Whitney U test.</p> <p>Learning Activities: Hands-on training using R, Seminars on application of the techniques</p>	CLO3

III 11 Hours	<p>Statistical Inference Confidence interval estimation - Classical Hypothesis Testing - General Principles; Testing for a mean: Two sample t-test; test on variance – F-test, ANOVA – One-way and Two-way Classifications, Association test-Fisher’s test - Chi-Square test, Statistical model. Tutorial.</p> <p>Learning Activities: Problem Solving on online available data, research paper presentation</p>	CLO4
IV 11 Hours	<p>Probability and Random Variables Introduction – Events - Conditional probability – Independence - Baye’s theorem; Random variables; Indicator random variables; Discrete probability distributions- A brief idea of normal, Poisson and binomial distribution.</p> <p>Learning Activities: Brainstorming, discussion and Problem Solving on random and online available data</p>	CLO5

Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Gupta S. C and Kapoor V. K.(2019). Fundamentals of Mathematical Statistics, 11/e. New Delhi: Sultan Chand and Sons publications
2. Daniel, W. W. and Cross C.L. (2018) Biostatistics: A Foundation for Analysis in the Health Sciences, 11/e. New York: John Wiley & Sons Inc.
3. Zar, J. H. (2010) Biostatistical Analysis, 5/e, Pearson Education.
4. Le, C. T., and L.E (2016). Eberly, Introductory Biostatistics, 2/e, Wiley sons.
5. Triola. M.M., M. F.Triola and J. Roy (2019). Biostatistics for the Biological and Health Sciences, 2/e, Pearson Education Ltd.
6. Motulsky, H, (2017). Intuitive Biostatistics: A Nonmathematical Guide to Statistical Thinking, 4/e. New York: Oxford University Press.
7. Gupta, S. C., & Kapoor, V. K. (2020). Fundamentals of mathematical statistics. Sultan Chand & Sons.
8. Grimmett, G., & Stirzaker, D. (2020). Probability and random processes. Oxford university press.

ONLINE MATERIAL

NPTEL

1. <https://nptel.ac.in/courses/111/106/111106112/>
2. <https://nptel.ac.in/courses/111/102/111102111/>
3. <https://nptel.ac.in/courses/102/106/102106051/>
4. <https://nptel.ac.in/courses/102/101/102101056/>

Course Title: Data Mining and Machine Learning Lab (Practical)**Course Code: BIM.518****Course type: SBC****Total Hours: 60**

L	T	P	Cr
0	0	4	2

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Perform dimensionality reduction to solve biological problems

CLO2: Build and evaluate regression models

CLO3: Handle large data by applying PCA and machine learning methods

CLO4: Apply classification as a tool to develop predictive platform

CLO5: Implement k-means and Hierarchical clustering procedure for the given biological data

Course Content

- (i). Applying dimensionality reduction analysis on given dataset: PCA, LDA using any programming language/tool
- (ii). How to generate Heat-Maps for RNA-Seq dataset
- (iii). Working with Hierarchical clustering on RNA-Seq data
- (iv). Performing linear regression analysis on training and estimating accuracy on testing set using any programming language/tool
- (v). Using k-Nearest Neighbors algorithm (K-NN) to classify the given dataset
- (vi). Using Random Forest to classify the given dataset
- (vii). Using Support Vector Machine (SVM) to make prediction for unknown dataset.
- (viii). Applying the concepts of Decision trees learning algorithm to predict the unknown target variable

Transactional Modes: Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Leskovec, J., Rajaraman, A. & Ullman, J. (2014). Mining of Massive Datasets.
2. Doupe, P., Faghmous, J., & Basu, S. (2019). Machine Learning for Health Services Researchers. Value in health: the journal of the International Society for Pharmacoeconomics and Outcomes Research, 22(7), 808–815.
3. Lan, K., Wang, D. T., Fong, S., Liu, L. S., Wong, K., & Dey, N. (2018). A Survey of Data Mining and Deep Learning in Bioinformatics. Journal of medical systems, 42(8), 139.
4. Bishop, C. (2007). Pattern Recognition and Machine Learning.

Course Title: Biostatistics using R Lab (Practical)

Course Code: BIM.519

Course type: SBC

Total Hours: 60

L	T	P	Cr
0	0	4	2

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Use R programming to analyse the biological data.

CLO2: Perform descriptive statistical analysis on a given data set

CLO3: Perform point and interval estimation of statistical parameters

CLO4: Fit appropriate probability distributions for a provided data set

Course Content

- (i). How to generate statistics summary (location, spread, shape, and peakedness)
- (ii). How to estimating data parameters: point and interval estimation- estimating mean, proportion, variance, variance ratio.
- (iii). Performing One sample and two Sample t tests on given dataset
- (iv). Performing F-test and ANOVA on given dataset
- (v). Performing the chi – square test of independence and Fisher's test.
- (vi). Practical understanding of Sampling distributions, Fitting of distributions (discrete and continuous)

Transactional Modes: Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Gupta S. C and Kapoor V. K.(2019). Fundamentals of Mathematical Statistics, 11/e. New Delhi: Sultan Chand and Sons publications
2. Daniel, W. W. and Cross C.L. (2018) Biostatistics: A Foundation for Analysis in the Health Sciences, 11/e. New York: John Wiley & Sons Inc.
3. Zar, J. H. (2010) Biostatistical Analysis, 5/e, Pearson Education.
4. Triola. M.M., M. F.Triola and J. Roy (2019). Biostatistics for the Biological and Health Sciences, 2/e, Pearson Education Ltd.
5. van Belle, G., L. D. Fisher, P. J. Heagerty, and T. Lumley (2004) Biostatistics: A Methodology for the Health Sciences, 2/e. John Wiley & Sons.

ONLINE MATERIAL

NPTEL

1. <https://nptel.ac.in/courses/111/106/111106112/>
2. <https://nptel.ac.in/courses/111/102/111102111/>

Course Title: Bioinformatics for Next Generation Sequencing Data

Course Code: BIM.520

Course type: VAC

Total Hours: 15

L	T	P	C
1	0	0	1

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Compare various NGS chemistries and NextGen Sequencing platforms

CLO2: Outline the complexity of NGS data, steps involved in NGS data analysis, and algorithms

CLO3: Describe data analysis approaches used for DNA and RNA sequencing using NGS platforms

CLO4: Examine the real-time application of NGS in medicine and agriculture using case studies

Units/ Hours	Contents	Mapping with CLO
I 4 Hours	Introduction to NGS techniques: Illumina sequencing, Roche 454 sequencing, Ion torrent, SOLiD sequencing, Nanopore sequencing. Learning Activities: Visit and demonstration of NGS platforms, Seminars on application of the techniques,	CLO1
II 3 Hours	Sequence quality evaluation - Phred Scores, Sequence file formats, Alignment of Next-Gen sequences to reference sequences Learning Activities: Hands-on training in sequence analysis, discussion and seminars on application of the techniques, Case studies	CLO2
III 4 Hours	NGS methods: Whole genome sequencing, Target sequencing, Exome sequencing, RNA seq, analysis- Transcriptome mapping, Methylation sequencing, ChIP-Seq. Learning Activities: Peer discussion, hands-on training in sequence analysis, research paper discussion	CLO3
IV 4 Hours	RNA-Seq data analysis, Differential Expression Analysis Workflow, Aligning Reads to Reference, Transcriptome Assembly, Applications of NGS Learning Activities: Peer discussion, hands-on training in sequence analysis, research paper discussion	CLO4

Transactional Modes: Lecture; Tutorial; Problem solving; Self-learning.

Suggested Readings

1. Brown, S. M. (2013). Next-Generation DNA Sequencing Informatics. Cold Spring Harbor Laboratory Press, 1/e.
2. Head, Steven R., Ordoukhanian, Phillip, Salomon, Daniel R (2018) Next Generation Sequencing Methods and Protocols, Humana Press Inc.
3. Xinkun Wang. (2016). Next-Generation Sequencing Data Analysis. Talyor and Francis Group.
4. Low, L., & Tammi, M. (2017). Introduction to next generation sequencing technologies. Bioinformatics: A Practical Handbook Of Next Generation Sequencing And Its Applications.
5. Lloyd Low and Martti Tammi (2017) A Practical Handbook of Next Generation Sequencing and Its Applications, World Scientific Publishing Co Pte Ltd
6. Beginner's Handbook of Next Generation Sequencing, Genohub
7. Shawn E. Levy and Richard M. Myers, "Advancements in Next-Generation Sequencing". Annu. Rev. Genom. Hum. Genet. 2016. 17:16.1–16.21

Course Title: Big Data Analytics in Healthcare

Course Code: BIM.521

Course type: DE

Total Hours: 45

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Discuss the fundamental concepts of big data

CLO2: Explain the methods used for large scale biological data handling

CLO3: Describe various approaches to generate protein big data

CLO4: Outline role of big data analytics in genomic research

CLO5: Explain the importance of big data analytics in healthcare

Units/ Hours	Contents	Mapping with CLO
I 11 Hours	<p>Fundamentals in Big Data Introduction to Big data and big data in bioinformatics. Techniques for handling big data- Data management, Data analytics. Map-Reduce Fundamentals, Technologies for handling big data- Vertical and Horizontal scaling techniques- Foundations of the Hadoop Ecosystem, Apache Spark</p> <p>Learning Activities: Group discussion, hands-on training in data handling, Peer discussion and student presentations</p>	CLO1 & CLO2
II 12 Hours	<p>Big Data Analytics in Protein Bioinformatics Protein Structure Alignment and Similarity Searching, functional assignment. Prediction of intrinsically disordered proteins using cloud computing. Mass spectrometry and NMR data analysis using inferential structure determination, low resolution crystallographic data analysis using probabilistic, Bayesian methods from SAXS and SANS data.</p> <p>Learning Activities: Hands-on training on biological data analysis, student presentations, research paper discussion</p>	CLO3
III 11 Hours	<p>Big Data Analytics in Genomics Challenges of Handling Genomic and Clinical Data, Big Data on the Cloud, Big data in NGS Read Alignment, Big Data Analytics in Calling Variants, Statistical Analysis of Genomic Data, Data mining using RNA seq data, Analysis of Genomic and Clinical Data. AI applications in clinical genomics</p>	CLO4

	Learning Activities: Hands-on training on biological data analysis, student presentations, research paper discussion	
IV 11 Hours	<p>Big Data Analytics in Health Research: Case studies on big data analytics for preventive and personalized medicine. Mining Massive Genomic Data for Therapeutic Biomarker Discovery in Cancer, Medical image processing and its role in healthcare data analysis. Big Data analytics-based models for various stages in healthcare.</p> <p>Learning Activities: Classroom presentation on real world application and peer discussion, Case studies, research paper discussion.</p>	CLO5

Transactional Modes: Lecture, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Mrozek, D. (2018). Scalable Big Data Analytics for Protein Bioinformatics. Springer Press.
2. Hurwitz, J. S., Nugent, A., Halper, F., & Kaufman, M. (2013). Big data for dummies. John Wiley & Sons.
3. Elmasri, R. (2021). Fundamentals of database systems seventh edition.
4. Lytras, M. D., & Papadopoulou, P. (Eds.). (2017). Applying big data analytics in bioinformatics and medicine. IGI Global.
5. Mrozek, D. (2018). Scalable big data analytics for protein bioinformatics. Computational Biology.
6. Li, S. (Ed.). (2020). Computational methods and data analysis for metabolomics. Totowa, NJ, USA:: Humana Press.
7. Wong, K. C. (2016). Big data analytics in genomics. Springer. Springer Press, 1st ed. 2016
8. Wong, K. C. (Ed.). (2016). Big data analytics in genomics. Springer.
9. Dey, N., Das, H., Naik, B., & Behera, H. S. (2019). Big data analytics for intelligent healthcare management. Academic Press. 1st ed.

Web resources

1. <https://doi.org/10.3390/ijms18020412>
2. <https://doi.org/10.1016/j.copbio.2019.03.004>
3. <https://doi.org/10.1007/s00521-019-04095-y>

NPTEL

1. https://onlinecourses.nptel.ac.in/noc20_bt10/preview
2. https://onlinecourses.nptel.ac.in/noc20_cs92/preview
3. https://onlinecourses.nptel.ac.in/noc22_bt20/preview

Course Title: Cheminformatics
Course Code: BIM.522
Course type: DE
Total Hours: 45

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Decode the molecular structure from various structural representations retrieved from the database

CLO2: Identify specific descriptors for small molecular compounds

CLO3: Develop a pharmacophore model from a set of drug molecule and quantify the structure activity relationship

CLO4: Virtual screening tools and efficiency assessments

Units/ Hours	Contents	Mapping with CLO
I 12 Hours	<p>Cheminformatics as a theoretical chemistry discipline: definition, main concepts and areas of application.</p> <p>Representing chemical structures on computers. Molecular graphs. Connectivity tables. Adjacency and distance matrices. Linear representations SMILES and SMIRKS. Hashed fingerprints. Exchange formats for chemical structures (MOL, SDF) and reactions (RXN et RDF). Chemical Databases. Different types of searching structures in the databases: exact match, sub-structural, super-structural and by similarity.</p> <p>Learning Activities: Classroom presentation and discussion on the topic, hands-on training in the chemical structure file formats and databases</p>	CLO1
II 11 Hours	<p>Molecular descriptors. Definition and main requirements. Different types of descriptors: constitutional, topological indices, geometry-based, surface based, substructural fragments, lipophilicity, etc. Development and validation of QSAR/QSPR models. Data preparation. Statistical parameters assessing models performance. Cross-validation. Models applicability domain. Ensemble modeling.</p> <p>Learning Activities: Problem based learning, Improved discussion, research paper discussion</p>	CLO2
III 11 Hours	<p>Molecular Interaction Fields. 3D QSAR. Molecular fields' similarity</p>	CLO3

	<p>Pharmacophore approach Pharmacophore features. Ligand- and structure-based pharmacophores. Merged and shared pharmacophores. Pharmacophore-based virtual screening</p> <p>Learning Activities: Peer discussion, Seminars on application of Pharmacophores, real world application, Problem Solving.</p>	
<p>IV 11 Hours</p>	<p>Chemical Space concept. Graph-based chemical space: scaffolds, frameworks and R-groups. Scaffold tree approach. Descriptor-based chemical space: distance and similarity metrics. Data visualization: Generative Topographic Mapping. Network-like similarity graphs. Activity landscapes. Bioisosteres. Activity cliffs. Virtual screening workflow. Drug-likeness filters and structural alerts. Parameters of screening efficiency.</p> <p>Learning Activities: small group projects, Case studies, research paper and peer discussion</p>	CLO4

Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Leach, A. R., & Gillet, V. J. (2017) .An Introduction to Cheminformatics, Springer,
2. Varnek, A.(2017). Tutorials in cheminformatics. John Wiley & Sons.
3. Engel, T., & Gasteiger, J. (2018). Cheminformatics: basic concepts and methods. John Wiley & Sons.
4. Sharma, N., Ojha, H., Raghav, P., & Goyal, R. K. (2021). Cheminformatics and Bioinformatics in the Pharmaceutical Sciences. Elsevier.
5. Stromgaard, K., Krogsgaard-Larsen, P., & Madsen, U. (2009). Textbook of drug design and discovery. CRC press. ISBN: 9780429111242.

Course Title: Molecular Evolution**Course Code: BIM.523****Course type: DE****Total Hours: 45**

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Describe evolutionary processes that give rise to variation in sequences and genomes

CLO2: Describe the architecture of the genome, contents and variation in base composition

CLO3: Outline the molecular level mechanisms of the various models in genome evolution

CLO4: Apply bioinformatics methods for studying genetic variation in and between species.

Units/ Hours	Contents	Mapping with CLO
I 12 Hours	Comparison of DNA sequences to calculate gene distance; Convergent and divergent evolution; Mutation Vs. Substitution-Rate of Molecular Evolution. Jukes Cantor Correction and evolutionary distance Learning Activities: Peer discussion, brainstorming, Case studies, research paper discussion.	CLO1
II 12 Hours	Genome evolution, RNA structure and evolution, Compensatory substitutions and the comparative method, Fitting evolutionary models to sequence data, The influence of thermodynamics on RNA sequence evolution Learning Activities: Classroom presentation and discussion on the topic, Problem based learning.	CLO2
III 10 Hours	Molecular clock- Concepts and significance-molecular mechanisms of molecular clock- Neutral theory -gene family organization. Applications of molecular phylogenetics Learning Activities: Discussion on each topic in a learner centric manner through term paper presentation, real world problems.	CLO3
IV 11 Hours	Paralogy and Orthology- coordination expression in evolution-genome: content, structure and evolution. Molecular evolution of recently diverged species - Databases of Molecular evolution. Learning Activities: Research paper presentation, Problem based learning sessions, Class quiz.	CLO4

Transactional Modes: Lecture, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Darwin, C.R. (1911). *On the origin of species by means of natural Selection, or preservation of favoured races in the struggle for life*. Hurst Publishers, UK.
2. Dawkins, R. (1996). *The Blind Watchmaker*, W.W. Norton & Company Jones and Bartlett Publishers.
3. Futuyma, D.J. (2009). *Evolution*. Sinauer Associates Inc. USA
4. Bromham, L. (2016). *An Introduction to Molecular Evolution and phylogenetics*. OUP Oxford.
5. Warnow, T. (2019). *Bioinformatics and Phylogenetics*. Springer International Publishing, ISBN 978-3-030-10836.

Web Resources

1. <http://www.bioinf.wits.ac.za/software/fire/evodb/>
2. <https://www.megasoftware.net/>

Semester III

Course Title: Research Methodology

Course Code: CCC.551

Course type: CC

Total Hours: 45

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to:

CLO1: Perform Literature survey, critically analyse the scientific problem and develop a research plan

CLO2: Use reference management systems and perform literature reviews using online resources

CLO3: Describe the importance of IPR and develops interest in entrepreneurship

CLO4: Write a good to technical report, manuscripts and scientific proposals

CLO5: Appreciate the importance of Research and Academic Integrity and follow safety protocols

Units/ Hours	Contents	Mapping with CLO
I 12 Hours	<p>Introduction: Meaning and importance of research, Different types and styles of research, Role of serendipity, Critical thinking, Creativity and innovation, Hypothesis formulation and development of research plan, Art of reading and understanding scientific papers, Literature survey, Interpretation of results and discussion.</p> <p>Learning Activities: Research paper presentation, Writing and Evaluation of research proposals, Peer discussion.</p>	CLO1
II 12 Hours	<p>Library: Classification systems, e-Library, Reference management, Web-based literature search engines, Intellectual property rights (IPRs).</p> <p>Learning Activities: Perform literature survey, Research paper presentation and group discussion.</p>	CLO2
	<p>Entrepreneurship and Business Development: Importance of entrepreneurship and its relevance in career growth, Types of enterprises and ownership.</p> <p>Learning Activities: Concept built with real examples, case studies, Student presentation and group discussion.</p>	CLO3
III 11 Hours	<p>Scientific and Technical Writing: Role and importance of communication, Effective oral and written communication, Scientific writing, Research paper writing, technical report writing, Making R and D proposals, Dissertation/Thesis writing, Letter writing and official correspondence, Oral</p>	CLO4

	<p>and poster presentation in meetings, Seminars, Group discussions, Use of modern aids; Making technical presentations.</p> <p>Learning Activities: Project report /research article preparation as a group activity, Research paper presentation</p>	
<p>IV 10 Hours</p>	<p>Research and Academic Integrity: Plagiarism, Copyright issues, Ethics in research, and case studies. Laboratory Safety Issues: Lab, Workshop, Electrical, Health and fire safety, Safe disposal of hazardous materials.</p> <p>Learning Activities: Case studies, Peer discussion, brainstorming, spontaneous quizzes</p>	<p>CLO5</p>

Transaction Mode: Lecture, demonstration, PPT.

Suggested Readings

1. Kumar, R. (2012). Research Methodology, SAGE Publications India Pvt. Ltd., New Delhi, India.
2. Gupta, S. (2005). Research Methodology and Statistical techniques, Deep and Deep Publications (P) Ltd. New Delhi, India.
3. Kothari, C.R. (2008). Research Methodology, New Age International, New Delhi, India.
4. Standard /Reputed Journal authors' instructions.
5. Denisova-Schmidt, E. (2021). Book Review: A Roadmap to the Future of Academic Integrity Research. Academy of Management Learning & Education.
6. Sutherland-Smith, W. (2008). Plagiarism, the Internet, and student learning: Improving academic integrity. Routledge.
7. Bretag, T. (Ed.). (2020). A research agenda for academic integrity. Edward Elgar Publishing.
8. Gould, J. R. (2020). Directions in Technical Writing and Communication. Routledge.

Course Title: Systems Biology
Course Code: BIM.524
Course type: CC
Total Hours: 45

L	T	P	Cr
2	0	2	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Relate the various biological pathways with consensus network motifs and therefore decipher the functioning

CLO2: Correlate the various biological pathways with consensus network motifs and therefore decipher the functioning

CLO3: Identify the optimal structure for analysing deep sequencing data and understand the main features of biological networks.

CLO4: Apply mathematical modelling to discuss relevant issues in Biology

Units/ Hours	Contents	Mapping with CLO
I 11 Hours	Introduction to system biology, networks, path, degree, cluster coefficient, scale-free networks and relevance, power law. Transcription networks, basic concepts, elements of Transcription networks, logic input functions. Learning Activities: Case study, research paper discussion, Quiz.	CLO1
II 12 Hours	Lac operon, Auto-regulation, a network motif: patterns, randomized networks and network motif, the feed forward loop (FFL) network motif, Case study in lactose system and arabinose system of <i>E. coli</i> , Dynamics of CI-FFL, IIFFL with logical gates, Case study of galactose utilization in <i>E. coli</i> , flagella system of <i>E.coli</i> , Convergent evolution of FFLs. Learning Activities: Problem Solving on various network study, case study, seminars on related topic	CLO2
III 12 Hours	Temporal programs and the global structure of transcription networks: SIM, BiFans, DORs, Interlocked FFL in <i>B. subtilis</i> sporulation network, Network motifs in developmental transcription networks: Network motif in PPI: Hybrid network motifs, hybrid FFL, Network motifs in neuronal networks: <i>C elegans</i> multi-input FFLs, Network motif in signal transduction networks. Learning Activities: Peer discussion, brainstorming discussion on related research papers.	CLO3
IV 10 Hours	Kinetic proofreading and conformational proofreading: genetic code, Precision of Translation, tRNA study, recognition of self and non-self by the immune system; Demand rule for gene regulation, optimal gene circuit design:	CLO4

	<p>cost of LacZ protein, fitness function and optimal expression level.</p> <p>Learning Activities: Problem Solving, Peer discussion and brainstorming quiz</p>	
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Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Ringrose, L. (2017). Epigenetics and Systems Biology, Elsevier Science.
2. Markus W. Covert (2017). Fundamentals of Systems Biology: From Synthetic Circuits to Whole-cell Models, CRC Press.
3. Chen, B. S., & Li, C. W. (2016). Big Mechanisms in Systems Biology: Big Data Mining, Network Modeling, and Genome-Wide Data Identification, Elsevier Science.
4. Klipp, E., Liebermeister, W., Wierling, C., & Kowald, A. (2016). Systems Biology, A Textbook, Wiley.
5. Alon, U. (2020). An Introduction to Systems Biology: Design Principles of Biological Circuits. Chapman & Hall, Second Edition.
6. Hake, S. and Wilt, F. (2003). Principles of Developmental Biology. W.W. Norton and Company, New York, USA.

Web resources:

1. The lac operon: <https://ocw.mit.edu/courses/biology/7-01sc-fundamentals-of-biology-fall-2011/molecular-biology/gene-regulation-and-the-lac-operon/#?w=535>

NPTEL

1. <https://nptel.ac.in/courses/102106035>
2. https://onlinecourses.nptel.ac.in/noc20_bt08/preview

MIT OpenCourseWare

1. <https://ocw.mit.edu/courses/biology/7-91j-foundations-of-computational-and-systems-biology-spring-2014/>

Course Title: Biological Database and Management System**Course Code: BIM.525****Course type: CC****Total Hours: 45**

L	T	P	Cr
3	0	0	3

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Apply the concept of RDBMS in understanding the organization of the biological data in various databases.

CLO2: Retrieve the data from various biological databases

CLO3: Use principles of DBMS to create solutions for biological problems.

CLO4: Develop SQL queries for the given scenario using DDL and DML commands

CLO5: Design and implement various control techniques in transaction processing system

Units/ Hours	Contents	Mapping with CLO
I 11 Hours	Introduction to Biological Databases: Nucleotide Sequence Databases, GenBank, DDBJ, EMBL, Sequence Flatfile and submission process, Protein sequence databases, Genomic databases, PDBsum, PDB, SCOP, CATH, Pathway and molecular interaction databases Learning Activities: practical using nucleotide/protein sequences, research paper discussion, data retrieval hand on training.	CLO1 & CLO2
II 12 Hours	Database planning and Design concepts General Database Planning and Design – Document or forms – preparation and architecture Entity- Relational ship Model- entities, Attributes, keys, tables design, relationships, roles and dependencies. Learning Activities: Peer discussion, research paper discussion.	CLO3
III 12 Hours	Relational DB Introduction to relational DB and transactions. SQL- statements-Data Definition-Manipulation-control-Objects, - Views, sequences and Synonyms. Working with code and forms- Front end development-query sublanguage-modifying relations in SQL. Learning Activities: Problem solving using codes, Debugging, Peer discussion, brainstorming and research paper discussion.	CLO4
IV	Transaction processing, concurrency control and recovery	CLO5

10 Hours	management. Transaction model properties, state serizability, lock base protocols, two phase locking. Learning Activities: Comparison debate, group discussion.	
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Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Negi, M.C. (2019). Fundamentals of Database Management System:Learn Essential Concepts of Database Systems. ISBN:9789388176620. BPB PUBN publications
2. Chopra, R. (2016). Database Management System (DBMS): A Practical Approach, 5th Edition. ISBN:9789385676345. S CHAND & Company Limited publications.
3. Ayyavaraiah, M., & Gopi, A. (2017). Database Management System.ISBN:9789386369703. Horizon Books (A Division of Ignited Minds Edutech P Ltd) publications
4. Date C.J. (2009). Introduction to database management. Vol1, Vol2, Vol3 addison Wesley.
5. Silberschatz, A., Korth, H. F., & Sudarshan, S (2005) Database system concepts. 5 Ed McGraw Hill Publications.
6. Elmasri, R., & Navathe, S. B. (2007). Fundamentals of Database systems. Benjamin Cummings Publishing Company. ISBN-10: 0321369572.
7. Rao, P. R. (2003). Database Management system, 3EdMcGraw Hill Publications. 9780071230575

Web Resources

1. <https://www.ncbi.nlm.nih.gov/>
2. expasy.org
3. <https://www.rcsb.org/>

Course Title: Biological Database and Management System Lab (Practical)**Course Code: BIM.526****Course type: SBC****Total Hours: 60**

L	T	P	Cr
0	0	4	2

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Apply principles of DBMS to create solutions for problems in biological sciences.

CLO2: Design an approach to create a Relational DBMS

CLO3: Define enforce integrity constraints on a database using a state-of-the-art RDBMS

CLO4: Create and query a database DBMS concepts

CLO5: Create non-redundant databases

Course Content

- (i). Creating and working with databases using MySQL, command to know the available database, Creating and Deleting a Database, Setting the Default Database
- (ii). Working with tables using MySQL: Creating and Deleting a Table - CREATE TABLE and DROP TABLE, adding content into the created table
- (iii). Working with MySQL codes on how to expand and edit the table: Inserting Rows, making use of insert, select, update and delete commands
- (iv). Codes for Querying the Database, making use of Comparison Operators, operator for String Pattern Matching, Arithmetic Operators, Logical Operators, querying by using different clause
- (v). How to produce a summary report using MySQL keywords, GROUP BY clause, GROUP BY Aggregate Functions and having clause
- (vi). Commands to modify and deleting data using MySQL: UPDATE-SET command, DELETE FROM command
- (vii). MySQL command to work with Nested Queries & Join Queries, Views, Functions [skip first/last] and Reports.
- (viii). Command to Work with more than one tables, setting keys (primary, foreign etc)

Transactional Modes: Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Negi, M.C. (2019). Fundamentals of Database Management System: Learn Essential Concepts of Database Systems. BPB PUBN publications
2. Chopra, R. (2016). Database Management System (DBMS): A Practical Approach, 5th Edition. CHAND & Company Limited publications.
3. Ayyavaraiah, M., & Gopi, A. (2017). Database Management System. Horizon Books (A Division of Ignited Minds Edutech P Ltd) publications
4. Silberschatz, A., Korth, H. F., & Sudarshan, S (2005) Database system concepts. 5 Ed McGraw Hill Publications.
5. Elmasri, R., & Navathe, S. B. (2007). Fundamentals of Database systems. Benjamin Cummings Publishing Company.
6. Rao, P. R. (2003). Database Management system, 3Ed McGraw Hill Publications. 9780071230575

Course Title: Biomolecular Simulations and Drug design Lab (Practical)**Paper Code: BIM.527****Course type: SBC****Total Hours: 60**

L	T	P	Cr
0	0	4	2

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Use the Linux environment for scientific computing

CLO2: Use the remote computing as a tool for high performance computation

CLO3: Use different energy minimization techniques

CLO4: Create macromolecular models from the amino acid sequence, and high-definition images using various graphics tools

CLO5: Use of different *in-silico* techniques for biomolecular simulations

CLO6: Perform molecular docking and high-throughput virtual screening.

Course Content

1. Linux basics and remote computing
2. Coordinate generations and inter-conversions of small molecules
3. Modelling macromolecular structure using homology modelling and *ab-initio* method.
4. Energy minimizations and optimization, *ab initio methods*
5. Advanced Visualization Software and 3D representations
6. Molecular Dynamics with GROMACS (a) Water structure and dynamics, (b) Binary Mixtures (c) HP36 in Water (d) Serotonin 1A in Membrane Bilayers
7. Review of Molecular Dynamics Principles
8. Molecular Recognition (a) Prediction of Protein-ligand interaction sites and (b) Prediction of Protein-protein interaction sites
9. Protein Ligand Docking using AutoDock
10. High throughput virtual screening using High throughput virtual screening using AutoDock Tools (ADT).

Transactional Modes: Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools.

Suggested Readings

1. Allen, M. P., & Tildesley, D. J. (2017). Computer Simulation of Liquids 2nd Edition, Oxford University Press.
2. Frenkel, D., & Smit, B. (2001). Understanding Molecular Simulation 2nd Edition, Academic Press.
3. Leach, A. R., & Leach, A. R. (2001)., Molecular Modelling Principles and Applications 2nd Edition. Pearson.
4. Alavi, S. (2020). Molecular Simulations: Fundamentals and Practice 1st Edition, Wiley-VCH.

Course Title: Entrepreneurship
Course Code: PCP.520
Course type: CF
Total Hours: 15

L	T	P	Cr
1	0	0	1

Course Learning Outcomes (CLO):

On completion of this course, students will be able to:

CLO1: Describe the nature of entrepreneurship and their role in economic development.

CLO2: Outline the behaviour and skills of an entrepreneur

CLO3: Explain the role of E-Cells in entrepreneurial development

CLO4: Write business plan/project proposals & managing start-up issues.

Units/ Hours	Contents	Mapping with CLO
I 4 Hours	The concept of entrepreneurship, the history of entrepreneurship. Entrepreneurial Structure; Nature, Characteristics, functions and its role in economic development. Entrepreneurship- problems and prospects in India Learning Activities: Students Teams, Student generated test questions, role playing	CLO1
II 4 Hours	Entrepreneurial behavior and skills, The entrepreneurial decision process, The skill gap analysis, role models, The entrepreneurial success stories Learning Activities: Case studies, Concept mapping, spontaneous quizzes	CLO2
III 3 Hours	Examination of emerging trends in the types of Computational sciences data being generated, The way it is managed and possibilities for new directions. Profiling the Computational entrepreneur. Learning Activities: Comparison debate, group discussion.	CLO3
IV 4 Hours	Funding opportunities for start-ups. Schemes of the Department of Science and Technology. Strategy and conduct of computational research. Understanding Business Models based on scientific computing and Managing Risk Learning Activities: Comparison debate, brainstorming, group discussion, student presentations.	CLO4

Transactional Modes: Videos and quizzes through the on-line LMS; Classroom learning (Videos, In-class Activities); Assignments and Projects; and Practical Experiences including challenges.

Suggested Readings

1. G. K. Varshney, (2012). Fundamentals of Entrepreneurship, Sahitya Bhawan Publications.
2. R. Roy, (2011). Entrepreneurship, 2nd Edition, Oxford
3. B.K. Mehta, (2018). Entrepreneurship and Small Business, SBPD Publishers.
4. Craig Shimasaki, (2020) Biotechnology Entrepreneurship Leading, Managing and Commercializing Innovative Technologies, 2nd Edition, Academic Press 2020
5. G.M. Javie, K. Li (2021) Chemistry Entrepreneurship, John Wiley & Sons, Inc.

Course Title: Enriching Bioinformatics
Course Code: BIM.528
Course type: DEC
Total Hours: 30

L	T	P	Cr
2	0	0	2

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Apply different skills to perform in competitive exams.

CLO2: Develop problem solving skills in the specific domain

CLO3: Revisit and revise the courses offered

CLO4: Solve exercises, mock tests and practice tests for competitive examinations

Units/ Hours	Contents	Mapping with CLO
I 8 Hours	Overview of replication, DNA damage and repair mechanisms, homologous and site-specific recombination. Learning Activities: Improved discussion, Problem based learning, discussion on most commonly used model organisms for genetic manipulation.	CLO1, CLO2, CLO3 & CLO4
II 7 Hours	Cell Signalling: Signalling through G-protein coupled receptors, signal transduction pathways. Learning Activities: Student seminars on application of the techniques in research and life, Peer group discussion on role of signaling pathways in research and health care.	
III 8 Hours	Zygote formation, cleavage, blastula formation, embryonic fields, gastrulation and formation of germ layers in animals. Learning Activities: Peer discussion on the existence of these developmental processes in different organisms, case studies, quiz.	
IV 7 Hours	Codominance, incomplete dominance, pleiotropy, sex linkage, sex limited and sex influenced characters. Learning Activities: Panel discussion on inherited diseases, Research paper presentation, Problem based learning sessions, Class quiz.	

Transactional Modes: Lectures; Problem solving; Self-learning.

Suggested Readings

1. Anthony, J.F., Miller, J.A., Suzuki, D.T., Richard, R.C., Gilbert, W.M. (1998). An introduction to Genetic Analysis. W.H. Freeman publication, USA.

2. Scott, F. and Gilbert, S.F. (2010). Developmental Biology. Sinauer Associates, Inc. USA.
3. Slack, J.M.W. (2005). Essential Developmental Biology, Wiley-Blackwell, USA.
4. Alberts, B., Bray, D., Lewis, J., Raff, M., Roberts, K. and Watson, J.D. (2010). Molecular Biology of the cell. Garland publishers, Oxford.
5. Celis, J.E. (2006). Cell biology: A laboratory handbook, Vol 1, 2, 3. Academic Press, UK.
6. Gupta, P.K. (2008). Cytology, Genetics and Evolution. Rastogi publications
7. Hancock, J. T. (2017). Cell signalling. Oxford University Press.
8. Prescott, D. (2012). Reproduction of eukaryotic cells. Elsevier.
9. Khatib, H. (2015). Molecular and quantitative animal genetics. John Wiley & Sons.

Web Resources

1. <https://www.embibe.com/exams/csir-net-question-paper/amp/>
2. <https://www.examrace.com/CSIR/CSIR-Sample-Old-Papers/Life-Sciences/>

Course Title: Dissertation Part-I
Course Code: BIM.600
Course type: SBC
Total Hours: 120

L	T	P	Cr
0	0	8	4

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Critically analyse, interpret, synthesize existing scientific knowledge based on literature review

CLO2: Demonstrate an understanding of the selected scientific problem and identify the knowledge gap

CLO3: Formulate a hypothesis and design an experimental/theoretical work

What Students would do:

Students will prepare a research proposal based on literature review and extensive student-mentor interactions involving regular discussions, meetings and presentations. Each student will submit a research/dissertation proposal of the research work planned for the M.Sc. dissertation with origin of the research problem, literature review, hypothesis, objectives, methodology to carry out the planned research work, expected outcomes and bibliography.

Students will have an option to carry out dissertation work in industry, national institutes or Universities in the top 100 NIRF ranking. Group dissertation may be opted, with a group consisting of a maximum of four students. These students may work using a single approach or multidisciplinary approach. Research projects can be taken up in collaboration with industry or in a group from within the discipline or across the discipline.

Evaluation Criteria:

The evaluation of the dissertation proposal will carry 50% weightage by supervisor and 50% by HoD and senior-most faculty of the department.

Dissertation Proposal (Third Semester)		
	Marks	Evaluation
Supervisor	50	Dissertation proposal and presentation
HoD and senior-most faculty of the department	50	Dissertation proposal and presentation

Modes of transaction: Group discussions and presentations; Self-Learning; Experimentation

Semester IV

Course Title: Dissertation Part-II

Course Code: BIM.601

Course type: SBC

Total Hours: 600

L	T	P	Cr
0	0	40	20

Course Learning Outcomes (CLO):

On completion of this course, students will be able to,

CLO1: Demonstrate an in-depth knowledge of scientific research pertaining to the area of study

CLO2: Demonstrate experimental/theoretical research capabilities based on rigorous hands-on training

CLO3: Critically analyse, interpret and present the data in light of existing scientific knowledge to arrive at specific conclusions

CLO4: Develop higher order thinking skills required for pursuing higher studies (Ph.D.)/research-oriented career options

Students will carry out their research work under the supervision of a faculty member. Students will interact with the supervisors through meetings and presentations on a regular basis. After completion of the research work, students will complete the dissertation under the guidance of the supervisor. The dissertation will include literature review, hypothesis, objectives, methodology, results, discussion, and bibliography.

Evaluation Criteria: The evaluation of dissertation in the fourth semester will be as follows: 50% weightage for continuous evaluation by the supervisor which includes regularity in work, mid-term evaluation, report of dissertation, presentation, and final viva-voce; 50% weightage based on average assessment scores by an external expert, HoD and senior-most faculty of the department. Distribution of marks will be based on report of dissertation (30%), presentation (10%), and final viva-voce (10%). The final viva-voce will be through offline or online mode.

Dissertation (Fourth Semester)		
	Marks	Evaluation
Supervisor	50	Continuous assessment (regularity in work, mid-term evaluation) dissertation report, presentation, final viva-voce
External expert, HoD and senior-most faculty of the department	50	Dissertation report (30), presentation (10), final viva-voce (10)

Transactional Modes: Lecture, Laboratory based Practical, Seminar, Group discussion, Team teaching, Self-learning, Online tools